



Fine Particulate Matter-induced Toxic Effects in an Animal Model of *Caenorhabditis elegans*

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ABSTRACT

Research has been focused on the health hazards of ambient PM_{2.5} related to humans. Many PM_{2.5} toxicity assessments using *in vitro* studies have focused on PM_{2.5}-bounded hazardous pollutants. However, PM_{2.5} toxicity assessment by *in vivo* studies allow for better observation of the overall effects of PM_{2.5} exposure on entire organisms, making *in vivo* PM_{2.5} toxicity assessment relevant. The toxic effects of outdoor PM_{2.5}, collected from National Pingtung University of Science and Technology (NPUST) and Linluo Junior High School (LJHS), Pingtung, Taiwan, on nematode *Caenorhabditis elegans* (*C. elegans*) were investigated. PM_{2.5} from NPUST and LJHS were found to be 4.5 and 2.5 μg Nm⁻³, respectively, which did not meet the standard. This levels of PM_{2.5} in Taiwan. For acute toxicity, no significant PM_{2.5} lethality on *C. elegans* was observed between NPUST and LJHS. PM_{2.5} from NPUST exhibited greater toxicity to lifespan (ageing), locomotion (head thrash), and reproduction (brood size) in the *C. elegans* animal models than that from LJHS; therefore, adverse effects could be correlated with PM_{2.5} concentrations. Prolonged exposure to PM_{2.5} led to more severe toxicity in nematodes as compared to acute exposure. In conclusion, this study suggests that the long-term adverse effects of ambient PM_{2.5} on environmental organisms should be carefully considered even when PM_{2.5} is at low levels. *C. elegans* is a sensitive animal model for the evaluation of PM_{2.5} ecotoxicity.

Keywords: *C. elegans*; PM_{2.5}; Lifespan; Locomotion; Reproduction; Ageing.

INTRODUCTION

Particulate matter 2.5 (PM_{2.5}) is a type of fine particles with an aerodynamic diameter of less than 2.5 μm. It is produced through processes including natural, indoor, and outdoor combustion processes, as well as other anthropogenic activities (Li *et al.*, 2018a; Martins and da Graça, 2018). Some of the identified major sources of PM_{2.5} emissions into the atmosphere are power plants (Mari *et al.*, 2016; Dodla *et al.*, 2017), waste incineration facilities (Yan *et al.*,

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2016), and industrial processes, such as the manufacture of steel stacks in the steelmaking industry (Gibson *et al.*, 2013; Owoade *et al.*, 2015). In addition, the large surface area of PM_{2.5} facilitates easy adhesion of toxic compounds such as polycyclic aromatic hydrocarbons (PAHs), polybrominated diphenyl ethers (PBDEs), polychlorinated biphenyls (PCBs), polychlorinated dibenzo-*p*-dioxins/furans (PCDD/Fs), on its surface (Pandey *et al.*, 2013; Chao *et al.*, 2016; Lee *et al.*, 2018). The World Health Organization (WHO) established guidelines on air quality that include limitations on exposure to PM_{2.5} of not more than 25 µg m⁻³ day⁻¹ and 10 µg m⁻³ year⁻¹ (Hopke *et al.*, 2018). In Taiwan, the allowable level of exposure to PM_{2.5} per year is 35 µg m⁻³, and for 24 hours, it is 15 µg m⁻³, as regulated by the Taiwan Air Quality Monitoring Network (TAQMN) of the Environmental Protection Administration, Taiwan (TEPA).

Due to the extremely fine size of PM_{2.5} particulates, they have been reported to be able to penetrate the blood brain barrier (Feng *et al.*, 2016; Lin *et al.*, 2017). Upon exposure, particulate matter induces overexpression of macrophages in the immune system and oxidative stress resulting in atherosclerotic plaques (Brook and Rajagopalan, 2010) and endothelial and vascular dysfunctions (Feng *et al.*, 2016). PM_{2.5} also affects the respiratory tract, particularly tiny sacs in the lungs called alveoli, where carbon dioxide and oxygen exchange takes place during respiration (Pinkerton *et al.*, 2000). This can further result in the activation of a series of inflammatory responses in the lungs (Chao *et al.*, 2018; Zhao *et al.*, 2019). PM_{2.5} is ubiquitous in the environment (Salako *et al.*, 2012; Mohammadyan *et al.*, 2017; Seneviratne *et al.*, 2017), and it has been reported that people suffering from cardiopulmonary and respiratory diseases are at high risk because their exposure to fine particles can cause short-term health effects such as eye, nose, throat and lung irritation, coughing, sneezing, runny nose, and shortness of breath (Kim *et al.*, 2018; Maciejczyk *et al.*, 2018; Polezer *et al.*, 2018). Several *in vivo* studies utilizing mice models have reported that exposure to airborne PM_{2.5} prolongs lung tumour progression (Yang and Xiao, 2018), promotes abdominal aortic aneurysms (Jun *et al.*, 2018), increases inflammatory cell infiltration (Hu *et al.*, 2017), and causes lipid accumulation and hepatic function loss (Xu *et al.*, 2019). *In vitro* studies have associated PM_{2.5} exposure to induced cytotoxicity and autophagy in human endothelial cells (Zhou *et al.*, 2018), as well as telomere lengthening (Miri *et al.*, 2019). Most importantly, epidemiological studies have shown that both short-term and long-term exposure to PM increases the risk of developing neurological disorders such as stroke (Fu *et al.*, 2019), Parkinson's disease (Hu *et al.*, 2019), and congenital heart disease as a result of prenatal exposure (Huang *et al.*, 2019), and generally stronger associations have been found with mortality and hospitalization (Karimi *et al.*, 2019). Recently, Lo *et al.* (2017) reported that PM_{2.5} was involved in deaths resulting from ischemic heart disease, stroke, lung cancer, and chronic obstructive pulmonary disease, wherein a fraction of 18.6% of the specified diseases was linked to PM_{2.5} in Taiwan. Therefore, PM_{2.5} is a major risk factor in Taiwan

(Lung *et al.*, 2016). Currently, there is no known established standard PM_{2.5} exposure level that may be referenced to avoid its adverse health effects (Kiesewetter *et al.*, 2015). Similarly, there is no singular *in vivo* animal model that can completely represent the human system. Therefore, there is a need for more research using other available model organisms to further understand the effects of PM_{2.5} on human health (Feng *et al.*, 2016).

Caenorhabditis elegans (*C. elegans*) was established by Sydney Brenner as a model organism in 1965 (Brenner, 1974). Compared with single-cell based *in vitro* assays, *C. elegans*-based assays allow for the evaluation of multiple toxicity endpoints (Kaletta and Hengartner, 2006). At the genetic level, more than half of the conserved signaling regions in humans are found in the nematode (Aoki and Mori, 2015). Numerous studies have reported the sensitivity of *C. elegans* in terms of pollution from different types of environmental media (Clavijo *et al.*, 2016; Zuo *et al.*, 2017; Rai *et al.*, 2019). However, only a few studies have reported the sensitivity of *C. elegans* to air pollutants, specifically, to PM_{2.5} (Zhao *et al.*, 2014; Sun *et al.*, 2015, 2016; Wang *et al.*, 2019). In a study by Sun *et al.* (2015), exposure to coal combustion-related PM_{2.5} induced lower response in all organism-level endpoints, specifically, prolonged exposure caused more severity than acute exposure to the nematodes. In addition, when Sun *et al.* (2016) determined the contribution of heavy metals found in coal combustion-related PM_{2.5}, he found that the combined exposure to specific metals caused greater toxicity than single exposure of said metals. Most importantly, transgenerational effects after exposure to diesel- and traffic-related PM_{2.5} were discovered by Wang *et al.* (2019) and Zhao *et al.* (2014) wherein reproductive disability had been transferred to later generations of *C. elegans*.

C. elegans is an inexpensive, bioethical, and easy model animal to maintain, with attractive physiological characteristics that can be used to assess the toxicological effects of environmental pollutants (Leung *et al.*, 2008). In this study, the toxicity of acute exposure to ambient outdoor PM_{2.5} samples collected from Southern Taiwanese schools such as National Pingtung University of Science and Technology (NPUST) and Linluo Junior High School (LJHS) was assessed using the different *C. elegans* toxicological endpoints, which include survival (lethality and lifespan), reproduction (brood size), developmental (growth), and locomotion (head thrash and body bend) endpoints.

MATERIALS AND METHODS

Chemical Reagents

The wild-type N2 strain of *C. elegans* was acquired from the Department of Biochemistry and Molecular Biology, College of Medicine, National Cheng Kung University (Tainan, Taiwan) and was maintained in nematode growth medium (NGM) plates containing bacteriological agar and bactopectone (Laboratories Conda, S.A., Spain) and NaCl (Honeywell Fluka™, New Jersey, USA). Additional NGM plate components such as CaCl₂, K₂HPO₄, and cholesterol were obtained from Sigma-Aldrich (St. Louis, MO, USA),

and MgSO_4 was acquired from Avantor Performance Materials, Ltd. (Gyeonggi-do, South Korea). OP50 *Escherichia coli* (*E. coli*) cultures were acquired from the Bioresources Collection and Research Center (Hsinchu, Taiwan) and Luria-Bertani broth was obtained from Sigma-Aldrich (St. Louis, MO, USA). For the bleaching solution, NaOCl was obtained from J.T. Baker (Central Valley, PA), and KOH was obtained from Duksan Pure Chemicals (Gyeonggi-do, South Korea). KH_2PO_4 used for the phosphate buffer was acquired from Avantor Performance Materials, LLC (Radnor, PA, USA), and Na_2HPO_4 used for the M9 buffer was obtained from Honeywell Fluka™ (New Jersey, USA). All physiological observations were done under a dissecting microscope (Olympus, SZX10, Waltham MA, USA). For the K-medium ingredients, NaCl was obtained from Honeywell Fluka™ (New Jersey, USA), and KCl was acquired from Avantor Performance Materials, Ltd. (Gyeonggi-do, South Korea). Dehydroethidium (DHE) for oxidative stress detection was purchased from Invitrogen™, Thermo Fisher Scientific (Waltham, MA USA).

Sample Collection

Outdoor ambient $\text{PM}_{2.5}$ samples were collected from two southern Taiwanese schools, NPUST and LJHS, located in Neipu, Pingtung County which is in the rural area. The air sampling site in NPUST was set up at the top floor of the College of Engineering in the NPUST campus. The other site in LJHS is nearby Linlou System Interchange of Formosa Highway. Each air sample was collected for 48 hours (2 days, $1 \text{ m}^3 \text{ min}^{-1}$) using a high-volume air sampler (SIBATA HV-1000R, Japan) following US EPA Reference Method TO9A. The $\text{PM}_{2.5}$ samples were collected onto quartz fibre filters that were pre-heated before sampling at 600°C for 2 hours. A balance with an accuracy of 0.1 mg was used to weigh the filter paper conditioned in an electronic desiccator before and after the sample collection for 24 h. The loaded filters were stored in a refrigerator at -20°C before extraction to limit the possible evaporation of volatile components.

Extraction, Purification, and Sample Preparation

After the collection of air samples, the mixture was extracted using a Soxhlet extractor with dichloromethane (DCM) for 24 h. The extracts were eluted with 15 mL DCM during the alumina column clean-up. The eluate was concentrated to approximately 1 mL and transferred to a vial. The concentrate was further concentrated to near dryness using a stream of nitrogen. For exposure concentrations, $\text{PM}_{2.5}$ was serially diluted 10X with 1% DMSO. The toxic effects in the study were limited in the organic fraction.

C. elegans Age Synchronization for Exposure Experiments

Wild-type N2 *C. elegans* were maintained on NGM plates seeded with *E. coli* as food and incubated at 22°C until the plates had a high density of eggs and gravid nematodes. These were then washed off the plates, placed into centrifuge tubes, and the gravid nematodes were lysed with a bleaching mixture leaving only the eggs to obtain age synchronized

populations of L1 nematode larvae in preparation for the exposure experiments and toxicity assays.

Acute Exposure

Synchronized L1 larvae were seeded onto NGM plates and incubated until L3 or young L4 stage. The L3 and young L4 worms were rinsed from the plate with K-medium, gently washed, and centrifuged at $2500 \times g$ for 4 minutes to remove the *E. coli* in the supernatant by aspirating without disturbing the nematode pellet. After washing, the pellet was re-suspended in K medium through continuous pipetting, and a seeding volume (worm per μL) was calculated by pipetting two 5 μL drops on a glass slide and manually counting the worms in each drop. Approximately 200 L3/young L4 worms were dispensed into each well of a 12-well plate each containing 1 mL of the different exposure concentrations for the two $\text{PM}_{2.5}$ samples (one set for NPUST and another for LJHS) diluted with K-medium. Acute (24-hour) exposure was performed at 22°C without the presence of food.

Lethality Assay

The lethality assay was performed by transferring 50 exposed nematodes (per exposure concentration) to NGM plates without food (OP50). These worms were gently prodded using the worm picker to assess viability. Nematodes that were non-responsive to stimulus were scored as dead. The assays were done in triplicates for each of the concentrations.

Lifespan Assay (Ageing Assay)

Fifty L3/young L4 worms from the acute exposure treatment were transferred to NGM plates with food (OP50). The worms were transferred to fresh NGM plates every day for the first 4 to 5 days (egg-laying period), and after that, the worms were no longer transferred. The plates were incubated at 22°C . Alive, censored (lost), and dead nematodes were recorded every 2 days within a span of 24 days (general lifespan of *C. elegans*). The survival plot or the Kaplan-Meier plot was constructed to evaluate the effects of the different concentrations of $\text{PM}_{2.5}$ on the lifespan or ageing of the nematodes. Triplicates were done for each of the exposure concentrations.

Reproductive or Brood Size Assay

L3/young L4 worms from the acute exposure treatment were assessed for 4 to 5 days of egg-laying. One L3/young L4 worm was transferred to each of the wells of a 12-well NGM plate with a fresh OP50 lawn. Each worm was transferred to a new plate every 2 days within the duration of egg-laying period. The old plates containing the eggs were hatched and incubated to L4 worms for easier counting of progeny. The total progeny for each of the worms was recorded. Twelve worms were evaluated for each of the exposure concentrations.

Growth Measurement Assay

L3/young L4 worms exposed to each of the $\text{PM}_{2.5}$ concentrations for 24 hours with prolonged exposure were

measured for their length to assess the effects of PM_{2.5} on the growth of the nematodes. Ten L3/young L4 nematodes exposed to each of the exposure concentrations were transferred to NGM plates with OP50 lawns and were incubated at 22°C for 48 hours until they reached the old L4 stage. The nematodes from each of the concentrations were exposed to a high temperature of 66°C in a dry bath for 5 minutes. This process straightens the body of the *C. elegans*, making it easier to measure the length. An optical microscope (Olympus) was used, and ImageJ software was utilized for the measurement of the worm length.

Locomotion Assay (Head Thrash and Body Bend Assay)

The endpoints of head thrash and body bending were used to evaluate the locomotion of the exposed L3/young L4 *C. elegans* by allowing the nematodes swim in K-medium. A head thrash was defined as a change in the direction of the mid body wherein the head and tail falls in the same direction as it moves. Body bending was regarded as a change in direction of the part of the nematodes corresponding to the posterior bulb of the pharynx along the y axis, assuming that the nematode was traveling along the x axis. Head thrashing was counted for 1 minute, and body bending was counted for 20 seconds. Thirty nematodes were examined per treatment, and three replicates were performed.

Statistical Analysis

The locomotion, brood size, and length measurement data for the nematodes did not meet the normal distribution using a normality test such as the Shapiro-Wilk test. The Kaplan-Meier plots for the survival rates were used to evaluate the effects of the different PM_{2.5} concentrations on the lifespan or ageing of the nematodes. The nonparametric Kruskal-Wallis *H* test was used to examine the differences between each the concentrations in comparison to the control. The survival plots or the Kaplan-Meier plot were derived for the lifespan data using GraphPad Prism 6 (San Diego, California, USA). The significance of each of the days were determined with the Kruskal-Wallis *H* test. Days

with significance were chosen and tested further. Each of the concentrations from the chosen days was also compared to the control using the Mann-Whitney *U* test. The statistical analyses were tested using SPSS version 12 (International Business Machines Corp., New York, USA).

RESULTS AND DISCUSSION

Lethality Effects of PM_{2.5} on *C. elegans*

Age-synchronized L4 worms were subjected to 24 hours of exposure to serially diluted samples of PM_{2.5}, after which lethality was investigated. No lethality was observed in any of the PM_{2.5} sample concentrations of NPUST (0.0014443–1444.3 mg L⁻¹) and LJHS (0.0011518–1151.8 mg L⁻¹) (Fig. 1) collected from NPUST and LJHS. Similarly, in a study conducted by Sun *et al.* (2015), coal combustion-related PM_{2.5} with exposure concentrations ranging from 0.01 to 100 mg L⁻¹ did not show any lethal effects on nematodes.

Life Span of *C. elegans* after PM_{2.5} Exposure

In this study, the toxicological endpoint lifespan was also evaluated since exposure of the nematodes to PM_{2.5} concentrations greater than 1 µg L⁻¹ was observed to cause toxicities. The concentrations of PM_{2.5} determined from NPUST and LJHS were 4.5 and 2.5 µg Nm⁻³ respectively. The detected levels of PM_{2.5} from both schools were lower than the allowable levels of outdoor PM_{2.5} in Taiwan, as regulated by the TAQMN. However, prolonged exposure showed that the lifespan of *C. elegans* was significantly reduced for all sample concentrations from the NPUST sampling site. Specifically, the lowest and highest concentrations (1.4443×10^{-3} mg L⁻¹ and 1.4443×10^3 mg L⁻¹) from NPUST caused the sharpest decrease in lifespan starting on post-exposure day 10 (Fig. 2). Additionally, a significant reduction in lifespan was observed on an earlier post-exposure day for LJHS for all concentrations. Concentrations of 1.5518×10^3 and 1.5518×10^2 mg L⁻¹ caused a rapid decrease in the population of the nematodes.

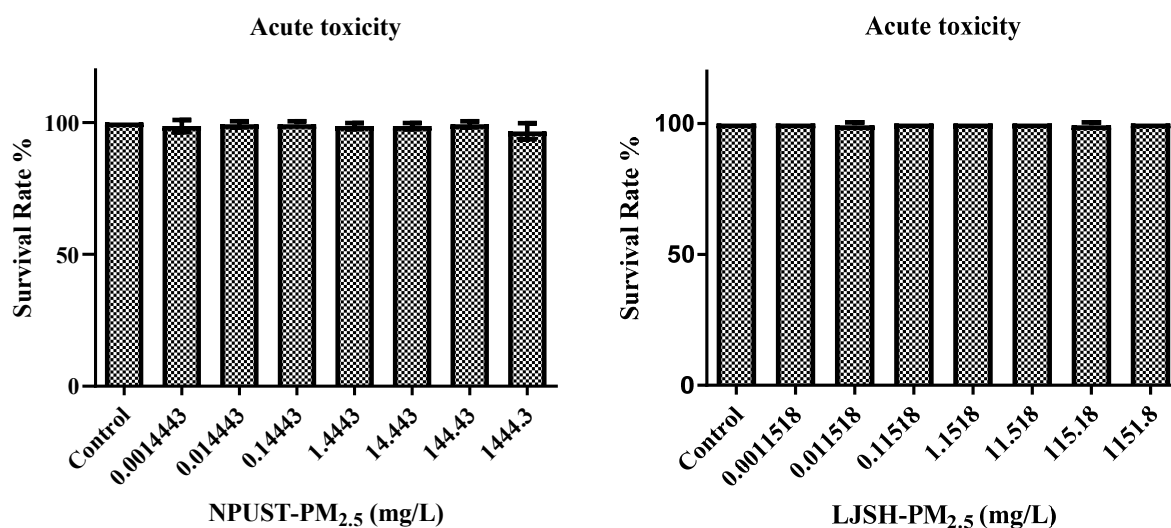


Fig. 1. Dose-dependent lethality of PM_{2.5} from (Left) NPUST and LJHS after 24-hour exposure.

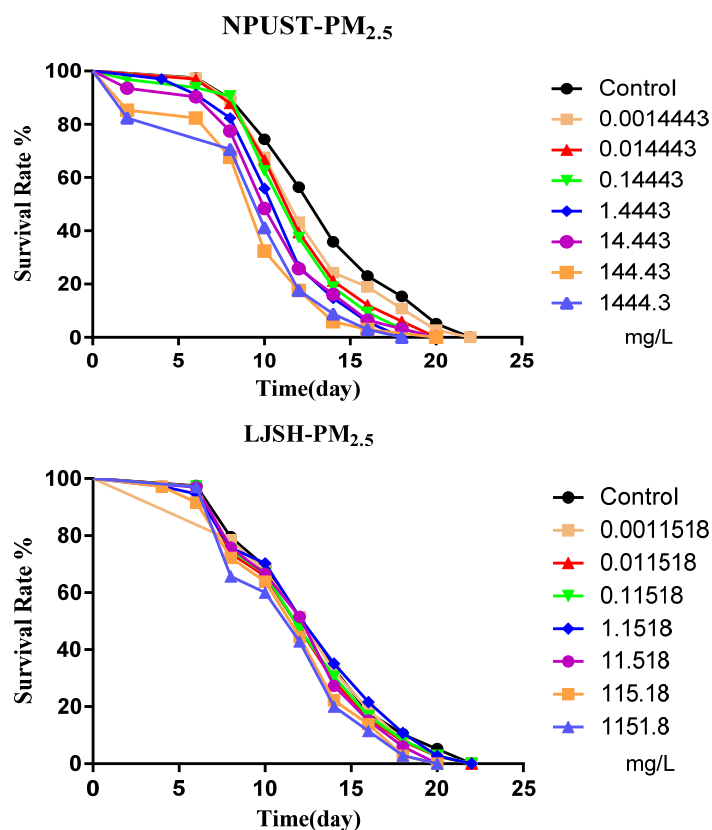


Fig. 2. Effects of PM_{2.5} from NPUST and (Bottom) LJHS on *C. elegans* lifespan after 24-hour exposure.

Lastly, *C. elegans* death occurred on post-exposure day 22, which is two days earlier than the usual lifespan of the nematode. In a similar report by Sun *et al.* (2015), coal combustion-related PM_{2.5} at 100 mg L⁻¹ significantly reduced the lifespan of *C. elegans*. However, there was no significant effect on the lifespan when the nematodes were exposed to concentrations 0.01–10 mg L⁻¹, which is inconsistent with the results of this study. Bases on the previous description of the Sun's study (Sun *et al.*, 2015), it was little concern for their study design. It was difficult to prevent from aggregation of PM_{2.5} after air sampling, but they didn't show and explain how to maintain PM_{2.5}-type particulate with diversely and homogenously widespread in the nematodes' medium. The finding of our studying is valuable due to our result indicating low levels of PM_{2.5} exposure in nematodes was still significantly linked into lower lifespan compared with the control. According to the previous reports and the present study (Zhao *et al.*, 2014; Sun *et al.*, 2015, 2016; Wang *et al.*, 2017), PM_{2.5} exposure decreased lifespan in the in-vivo and epidemiological studies including coal combustion in *C. elegans* (Sun *et al.*, 2015, 2016), traffic-related pollutants in the *C. elegans* (Zhao *et al.*, 2014), ambient air in *Drosophila* (Wang *et al.*, 2017), ambient air in newborns (Martens *et al.*, 2017), and ambient air in elderly (Li *et al.*, 2018b). Therefore, the level of outdoor air PM_{2.5} from the two schools, although lower than the national standard in Taiwan, was still shown to have a significant impact on microorganisms and ultimately may pose a threat on human health. To this end, additional

endpoints (such as reproductive and locomotion) were investigated to further confirm this effect.

Growth Measurement Assay

No lethality was observed when the nematodes were subjected to acute exposure with the different concentrations of PM_{2.5} from either sampling site (NPUST and LJHS). Also, specified concentrations of samples from both the NPUST (1.4443×10^{-3} to 1.4443×10^3 mg L⁻¹) and LJHS (1.5518×10^{-3} to 1.5518×10^3 mg L⁻¹) sampling sites did not lead to the alterations in body length (Fig. 3). Results showed that in all concentrations, the body length of the nematodes remained at 1 mm, the standard length of *C. elegans* (Leung *et al.*, 2008). A similar study by Sun *et al.* (2015) reported that acute and prolonged exposure of *C. elegans* to PM_{2.5} of concentrations 0.01 to 10 mg L⁻¹ had no significant effect on the body lengths of the nematodes under study; however, acute exposure to PM_{2.5} at a concentration of 100 mg L⁻¹ and prolonged exposure to PM_{2.5} at concentrations of 10 to 100 mg L⁻¹ significantly reduced body length. Our result was inconsistent with that discussed in Sun's study (Sun *et al.*, 2015).

Effect of Outdoor Air PM_{2.5} Levels on the Reproductive System of *C. elegans*

The secondary targeted organs of environmental pollutants include the neurons and the reproductive system (Leung *et al.*, 2008; Aoki and Mori, 2015). In the *C. elegans* model, the reproductive system is one of the major targets of

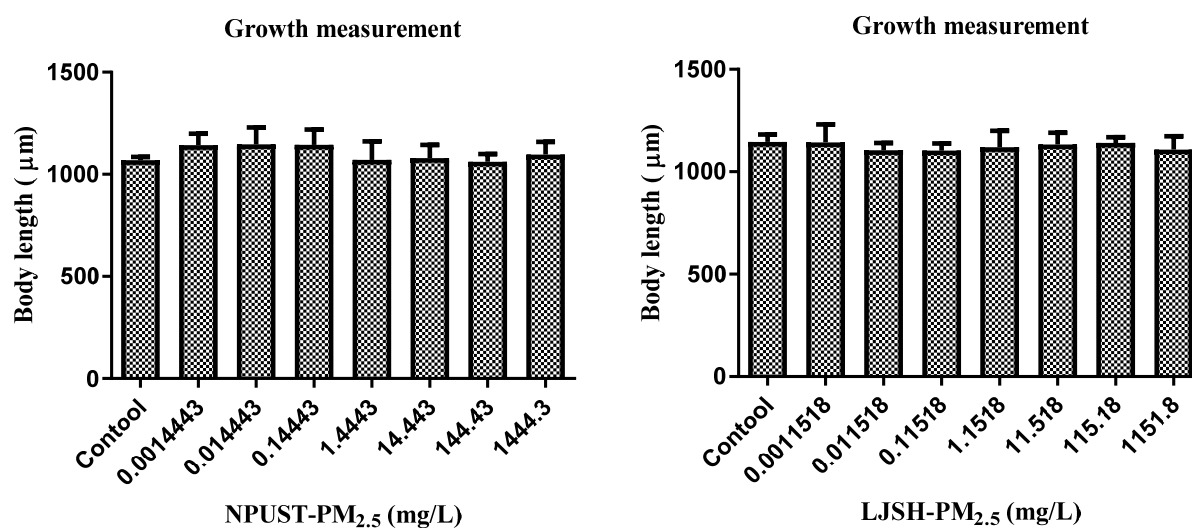


Fig. 3. Effects of PM_{2.5} from (Left) NPUST and LJHS on *C. elegans* growth, observed as changes in body length, after 24-hour exposure.

toxicants in the body, next to the neurosensory system (Aoki and Mori, 2015). In order to understand the effects of PM_{2.5} on its translocation in *C. elegans*, the brood size was also assessed for each of the worms exposed to different PM_{2.5} concentrations (Fig. 4). The total number of eggs, or brood size, was determined twice per concentration of samples from NPUST and LJHS. After acute exposure, the first measurement (Fig. 4(A)) showed that brood size was significantly reduced as the concentration of samples from NPUST increased. Specifically, brood size at $1.4443 \times 10^{-3} \text{ mg L}^{-1}$ was reduced to 18 eggs laid per worm, and concentrations from 1.4443×10^{-2} to $1.4443 \times 10^3 \text{ mg L}^{-1}$ reduced the brood size to 21, 11, 10, 10, 10, and 3 eggs laid per worm, respectively. Similarly, the second assessment of brood size (Fig. 4(B)) showed that at concentrations ranging from 1.4443×10^{-3} to $1.4443 \times 10^3 \text{ mg L}^{-1}$, the eggs laid per worm decreased to 13, 6, 5, 2, 6, 3, and 1, respectively. For samples gathered from LJHS, the first and second assessment of brood size (Figs. 4(C) and 4(D)) showed no significantly different number of eggs laid per worm at concentrations ranging from 1.5518×10^{-3} to $1.5518 \times 10^3 \text{ mg L}^{-1}$. In this study, the results showed that exposure to high ambient PM_{2.5} concentrations had a significant effect on the reduction of brood size of *C. elegans*. This result is analogous to the study conducted by Wang *et al.* (2019), wherein diesel particulate matter (DPM) caused reproduction deficits in *C. elegans*, as shown by a decrease in brood size during the first generation of offspring. Moreover, in the same study, it was observed that consecutive generation exposure to DPM led to severe limitations in brood size.

Currently, studies have shown that the self-progeny of *C. elegans* is an effective parameter that can be used to determine the extent of reproductive disability due to the toxicity of various substances (Wang *et al.*, 2018; Moon *et al.*, 2019). In general, the effect can be observed in a dose-dependent manner (Yang *et al.*, 2018). This was not the case for this study. The control group generated a relatively low brood size of 18. Dietary restriction (DR) can

significantly lower the progeny number in *C. elegans* (El-Hajj and Newman, 2015). In addition, for the size, PM_{2.5} is greater than *E. coli* OP50 ($2.5 \text{ µm} > 2 \text{ µm}$, general food source of *C. elegans*) (Khan *et al.*, 2018). Therefore, toxicity of PM_{2.5} is related to the size of the exposed organism.

Effects of Ambient PM_{2.5} on Locomotion Behavior of *C. elegans*

Locomotion assays used to analyze the head thrashing and body bending movements of the nematodes were conducted to assess the neurological toxicity brought about by their exposure to different concentrations of PM_{2.5} (Fig. 5). A significant reduction in head thrashing movement and body bending were observed for nematodes exposed to PM_{2.5} sampled from NPUST as compared to nematodes exposed to PM_{2.5} sampled from LJHS. These results imply that levels of PM_{2.5} may be correlated to the greatly disrupted functions of the reproductive and neurosensory systems in *C. elegans*. Additionally, prolonged exposure to PM_{2.5} was observed to cause more sensitivity to *C. elegans*. Zhao *et al.* (2014) established that acute exposure to high concentrations of PM_{2.5} causes adverse effects on both the reproduction and neurosensory functions of *C. elegans*. Moreover, prolonged exposure to low concentrations of PM_{2.5} further significantly decreased the locomotion behavior of the nematodes. However, this was true for exposure to pollutant concentrations in the range of $\mu\text{g L}^{-1}$. This is because of the greater dispersion of PM_{2.5} in DMSO, which increased the bioavailability of the particulate matter to the nematodes (Artifon *et al.*, 2019). The main route of uptake of pollutants in *C. elegans* is through the alimentary system; the worms actively ingest particulates while feeding on bacteria. The presence of bacterial food causes the adsorption of a proportion of the particles and enhances the oral uptake, resulting in higher toxicity (Gonzalez-Moragas *et al.*, 2015). Therefore, the dispersion of particulate matter in the media of choice is important in assessing their effects on *in vivo* systems.

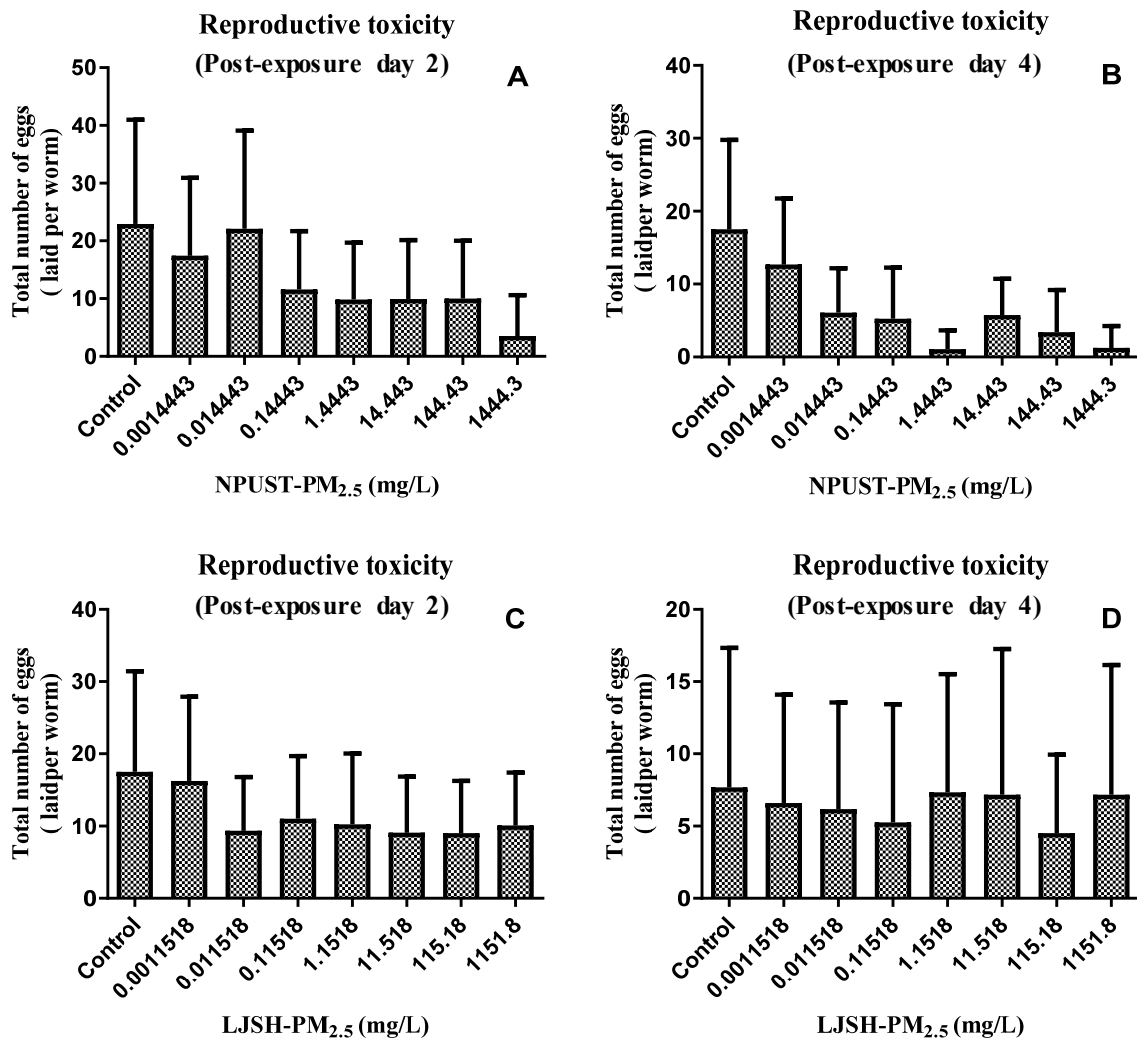


Fig. 4. Effects of PM_{2.5} from NPUST and (Bottom) LJHS on *C. elegans* brood size, observed (A and C) two days and (B and D) four days after 24-hour acute exposure.

Several toxic endpoints were examined in the present study. Low PM_{2.5} levels were found in our sampling areas. To make our amazement, the low levels of PM_{2.5}-bounded pollutants induced several adverse effects including accelerated aging process, reduced reproduction, and delayed locomotion. The present study was limited based on the air sampling technique and pretreatment protocol. The standard methods of PM_{2.5} air sampling and PM pretreatment were used in our study. This study excluded in the toxicity in inorganic fraction due to use of the Soxhlet extractors. The present study only considered the toxic effects of PM_{2.5}-bounded hazardous pollutants after the high-volume PM_{2.5} air samplers mainly due to occurrence of aggregation during the air sampling. Future work will be encouraged to gather airborne PM_{2.5}-type particulate directly into nematodes' medium before aggregation.

CONCLUSIONS

Low PM_{2.5} levels were obtained at NPUST and LJHS in this study. It was found that ambient PM_{2.5} levels at NPUST

were higher than the PM_{2.5} level at LJHS. In addition, the *in-vivo* model of *C. elegans* was utilized to evaluate the possible health risks associated with differing PM_{2.5} levels through the investigation of multiple toxic endpoints namely, lethality, lifespan, development, locomotion, and reproduction. The acute toxicity and growth measurement of *C. elegans* was unaffected by PM_{2.5} collected from NPUST and LJHS, but its lifespan was significantly reduced in the case of airborne PM_{2.5} at NPUST starting on post-exposure day 10. For the reproductive toxicity and locomotion, reduced brood size and the frequency of head thrashes for NPUST-PM_{2.5} was observed, but there were no significant differences in lifespan, brood size, head thrashes, and body bends between the control and LJHS-PM_{2.5}. Our significant findings were to indicate the potential toxic effects of long-term exposure to low PM_{2.5} on *C. elegans* models, even though the measurements were lower than PM_{2.5} standards. This study implored that the long-term adverse effects of ambient PM_{2.5} on environmental organisms should be carefully considered even though PM_{2.5} levels didn't meet the standards.

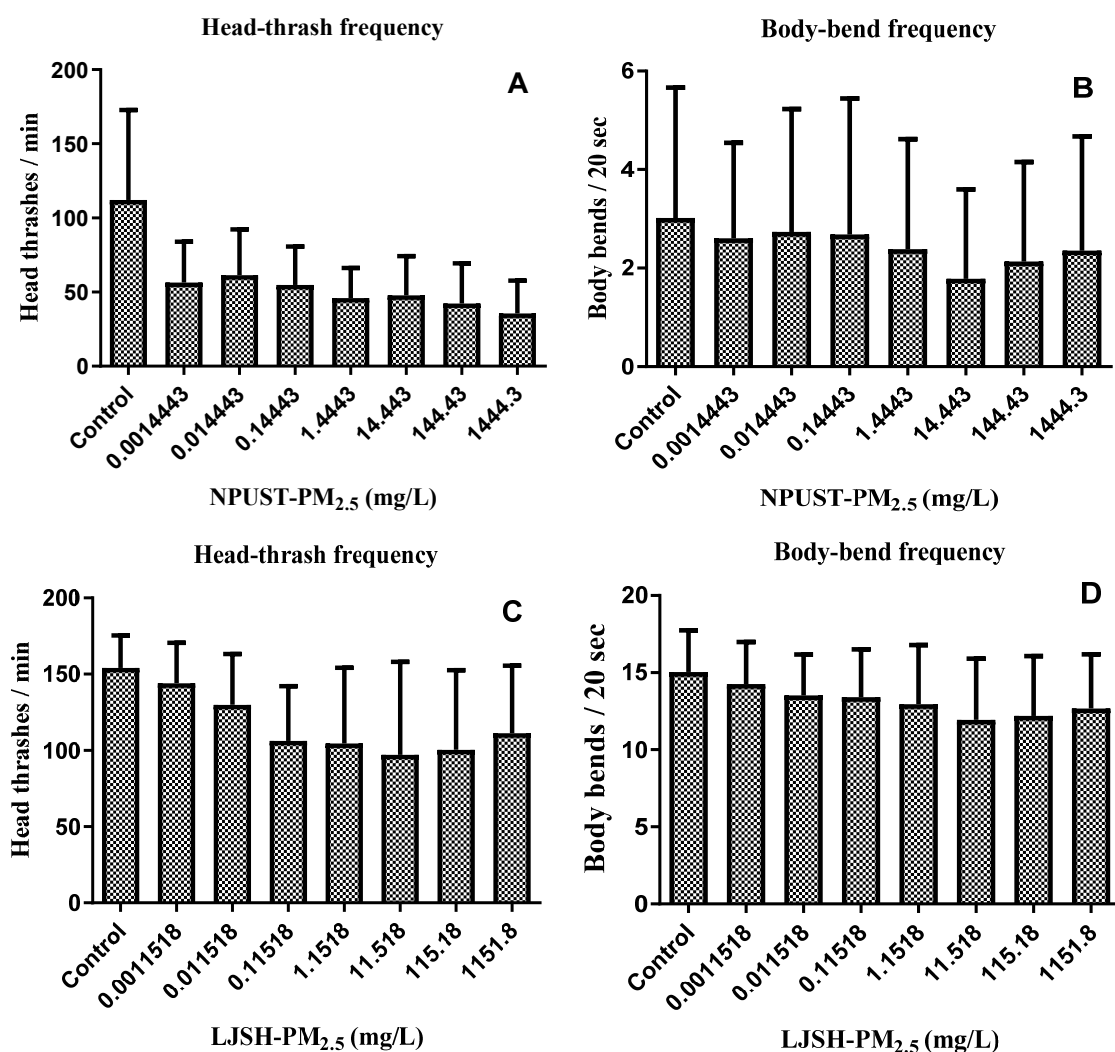


Fig. 5. Effects of PM_{2.5} from NPUST and (Bottom) LJHS on *C. elegans* locomotion and movement after 24-hour acute exposure. A and C for head thrashing and B and D for body bending.

ACKNOWLEDGMENTS

This study was supported by a grant from the Ministry of Science and Technology (MOST 106-2221-E-020-001-MY3). We acknowledge Mr. Ying-Jih Syu from National Pingtung University of Science and Technology for assisting us to maintain and culture *C. elegans*. We also want to thank Miss Chia-Jung Yen from Kaohsiung Medical University for assisting us with the *C. elegans* experiments. We would also like to thank Dr. Chang-Shi Chen at National Cheng Kung University for his advice and help in attaining the *C. elegans* culture. The authors thanked the teachers and students in Linluo Junior High School for providing the air sampling site.

DISCLAIMER

The authors declare no conflicts of interest.

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Received for review, March 21, 2019

Revised, April 23, 2019

Accepted, April 24, 2019